

## Product Information

# CellLytic™ MEM Protein Extraction Kit

**CE0050**

## Product Description

The CellLytic™ MEM Protein Extraction Kit offers a fast and convenient method to isolate hydrophobic and raft-associated proteins (proteins associated with glycosphingolipids and cholesterol rich membrane microdomains) from cells. The method, based on phase separation, does not require cell membrane isolation. Hydrophobic and raft-associated proteins are concentrated in the hydrophobic phase and may be used for further experiments. The CE0050 kit may be used in conjunction with the following techniques:

- Immunoprecipitation
- Gel electrophoresis (Coomassie® Brilliant Blue staining, and silver staining)
- Dot blot and Western blot

The kit has been tested with the following cell lines: HeLa, HEK-293, NIH 3T3, COS, and CHO.

Several publications<sup>1-6</sup> and dissertations<sup>7-10</sup> have cited use of product CE0050 in their protocols.

## Precautions and Disclaimer

For R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

## Components

- Lysis and Separation Buffer (Component L2417): 50 mL
- Wash Buffer for CellLytic™ Kit (Component W2514): 50 mL
- Sodium Chloride, 4 M solution (Component S2568): 1.5 mL
- Protease Inhibitor Cocktail for mammalian cell and tissue extracts (Component P8340): 1 mL

## Reagents and Equipment Required

(Not provided)

- Tissue culture centrifuge tubes, 15 mL conical
- Microcentrifuge tubes
- Serological centrifuge
- Microcentrifuge, refrigerated
- Dulbecco's Phosphate Buffered Saline (DPBS, such as Catalogue Number D8537)
- PBS containing 1 mM EDTA
- Cell scraper (such as Catalogue Number CLS3010)

## Storage/Stability

CE0050 ships on cooler packs ("wet ice"). It is recommended to store the unopened kit at -20 °C. After opening the kit:

- Store Component P8340 (Protease Inhibitor Cocktail for mammalian cell and tissue extracts) at -20 °C.
- Store the other Components at 2-8 °C.

## Preparation Instructions

### Preparation of the Lysis and Separation Working Solution

- Always store the Lysis and Separation Buffer (L2417) refrigerated, or on ice.
- Mix the Lysis and Separation Buffer before use.
- Prepare the Lysis and Separation Working Solution fresh daily, by adding 6 µL of P8340 to 600 µL of L2417.

## Modifications of the Lysis and Separation Working Solution and the Wash Buffer

- The Lysis and Separation Buffer can be diluted with the Wash Buffer (1:1), to decrease the volume of the hydrophobic phase. This will result in a more concentrated hydrophobic protein sample.
- The Lysis and Separation Working Solution, prepared with Lysis and Separation Buffer diluted with the Wash Buffer, does not significantly affect the protein yield in the hydrophobic phase.
- The Lysis and Separation Buffer and the Wash Buffer contain 150 mM NaCl.
  - Some hydrophobic proteins require a higher salt concentration for extraction into the hydrophobic phase.
  - Use the supplied 4 M NaCl solution to adjust the NaCl concentration to the required level for the protein(s) of interest.

## Procedure

This procedure is suitable for extraction of  $10^6$  to  $10^7$  cells. If larger numbers of cells are to be used, or multiple extractions are run in parallel, adjust the procedure accordingly.

### Cell collection

For adherent cells, Steps 1-7 are required.

For cells growing in suspension, begin with Step 5.

1. Aspirate the growth medium from the tissue culture vessel.
2. Wash the cells with Dulbecco's PBS.
3. Add 0.1 mL of PBS containing 1 mM EDTA solution per each  $\text{cm}^2$  of culture area.
4. Incubate until the cells detach. (Alternatively, add PBS, and then scrape the cells using a cell scraper.)
5. Transfer the cell suspension into a 15 mL conical tube.
6. Collect the cells by centrifugation at  $600 \times g$  for 5 minutes.
7. Aspirate the supernatant. Store the pellet on ice.

## Separation of hydrophobic and hydrophilic proteins

1. Mix the prepared Lysis and Separation Working Solution containing the Protease Inhibitor Cocktail before use.
2. Resuspend  $10^6$ – $10^7$  cells in 600  $\mu\text{L}$  of ice-cold Lysis and Separation Working Solution.
3. Mix gently by pipetting up and down. Vortex briefly.
4. Transfer the suspension to a microcentrifuge tube.
5. Incubate the cell suspension on ice for 10 minutes.
6. Centrifuge the cell lysate in a pre-cooled ( $4\text{ }^\circ\text{C}$ ) microcentrifuge at  $10,000 \times g$  for 5 minutes.
7. Transfer the clarified lysate to a new microcentrifuge tube. A 30-50  $\mu\text{L}$  aliquot of the total protein lysate may be saved for further analysis.
8. Incubate the lysate at  $30\text{ }^\circ\text{C}$  for 3-5 minutes. During the incubation, mix once by inverting the tube. The lysate will turn cloudy during the incubation. (**Note:** Incubation at  $30\text{ }^\circ\text{C}$  is preferable. However, incubation at temperatures up to  $37\text{ }^\circ\text{C}$  is possible.)
9. Centrifuge the tube in a microcentrifuge at room temperature at  $3000 \times g$  for 3 minutes. Ensure the centrifuge temperature is higher than  $20\text{ }^\circ\text{C}$ . **Do not return the tube to ice after centrifugation.**
10. Transfer the upper hydrophilic phase containing hydrophilic proteins to a new microcentrifuge tube. The lower hydrophobic phase is greatly enriched with hydrophobic and raft-associated proteins.
11. To remove residual hydrophilic proteins from the hydrophobic phase, the hydrophobic phase may be washed with the Wash Buffer:
  - Add 400  $\mu\text{L}$  of the Wash Buffer to the hydrophobic phase.
  - Mix gently.
  - Incubate the tube on ice for 10 minutes.
  - Repeat Steps 9-10.

## Downstream applications

### SDS-PAGE electrophoresis

Samples from the hydrophilic and hydrophobic phases can be used directly for acrylamide gel electrophoresis. The dye front may appear diffuse, but the final protein pattern is not affected. However, some PAGE systems will require the samples to be diluted 5-10 fold in order to obtain good resolution.

Alternatively, the samples can be precipitated with TCA, to obtain more concentrated samples.

For comparative analysis of protein separation between the phases, it is recommended to normalize the samples loaded on the gel.

### Dot blot

For fast analysis of an extraction for a specific protein, a dot blot can be performed using 1-2 µL samples.

### Immunoprecipitation

Before adding the immobilized antibody, dilute the hydrophobic phase 10-fold to 12-fold with Wash Buffer, to make the solution compatible with antibody binding.

### Other applications

For applications requiring low salt concentrations, the sample may be dialyzed.

## References

1. Kami, D. *et al.*, *BMC Biotechnol.*, **13**, 102 (2013).
2. Kong, X. *et al.*, *Mol. Biol. Cell*, **27(25)**, 3972-3979 (2016).
3. Morelle, W. *et al.*, *J. Clin. Endocrinol. Metab.*, **102(4)**, 1375-1386 (2017).
4. Kumar, S.V. *et al.*, *Sci. Rep.*, **10(1)**, 14985 (2020).
5. Matsuda, A. *et al.*, *J. Proteome Res.*, **19(6)**, 2516-2524 (2020).
6. van Tilburg, A.Y. *et al.*, *Microb. Biotechnol.*, **15(5)**, 1633-1651 (2022).
7. Kumar, Sushmitha Vijaya, "Investigating the Role of Carotenoids in Membrane Organization of *Pantoea* sp. YR343". University of Tennessee Knoxville, Ph.D. dissertation, p. 69 (2018).

8. Mielich-Süß, Benjamin, "Elucidating structural and functional aspects of prokaryotic membrane microdomains". Julius-Maximilians-Universität Würzburg, Ph.D. dissertation, pp. 85, 107 (2017).
9. Potelle, Sven, "TMEM165: un nouvel acteur de la regulation de l'homéostasie golgienne du Mn<sup>2+</sup>, impliqué dans les anomalies congénitales de la glycosylation" ("TMEM165: a new regulator of Golgi Mn<sup>2+</sup> homeostasis involved in Congenital Disorders of Glycosylation"). Université de Lille Nord de France - Lille I, Ph.D. dissertation, p. 129 (2017).
10. Burdiel, Laura Cueto, "The effects of the synthesis and localization of ZipA, an essential component of the *Escherichia coli* divisome, on division and membrane dynamics ". Universidad Autónoma de Madrid, Ph.D. dissertation, p. 57 (2018).

## Notice

We provide information and advice to our customers on application technologies and regulatory matters to the best of our knowledge and ability, but without obligation or liability. Existing laws and regulations are to be observed in all cases by our customers. This also applies in respect to any rights of third parties. Our information and advice do not relieve our customers of their own responsibility for checking the suitability of our products for the envisaged purpose.

The information in this document is subject to change without notice and should not be construed as a commitment by the manufacturing or selling entity, or an affiliate. We assume no responsibility for any errors that may appear in this document.

## Technical Assistance

Visit the tech service page at [SigmaAldrich.com/techservice](https://www.sigmaaldrich.com/techservice).

## Terms and Conditions of Sale

Warranty, use restrictions, and other conditions of sale may be found at [SigmaAldrich.com/terms](https://www.sigmaaldrich.com/terms).

## Contact Information

For the location of the office nearest you, go to [SigmaAldrich.com/offices](https://www.sigmaaldrich.com/offices).

The life science business of Merck KGaA, Darmstadt, Germany operates as MilliporeSigma in the U.S. and Canada.

MilliporeSigma, and Sigma-Aldrich are trademarks of Merck KGaA, Darmstadt, Germany or its affiliates. All other trademarks are the property of their respective owners. Detailed information on trademarks is available via publicly accessible resources.

© 2022 Merck KGaA, Darmstadt, Germany and/or its affiliates. All Rights Reserved.

CE0050pis Rev 07/22 EM,PY,CMH,GCY,MAM

**MILLIPORE  
SIGMA**