

# Restriction Endonuclease Alu I

From Arthrobacter luteus

Cat. No. 10 239 275 001 Cat. No. 10 656 267 001

500 units (10 U/μl) 2000 units (10 U/μl)



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Store at -15 to  $-25^{\circ}$ C

Stability/Storage

The undiluted enzyme solution is stable when stored at -15 to -25°C until the expiration date printed on the label. Do not store below -25°C to avoid freezing.

Sequence specificity Alu I recognizes the sequence AG/CT and generates fragments with blunt termini (1).

Compatible ends

The enzyme generates compatible ends to any blunt end.

Isoschizomers

Alu I is not known to have isoschizomers

Methylation sensitivity

Alu I is inhibited by the presence of 6-methyladenine, 5-methylcytosine, 5-hydroxymethylcytosine or 4-methylcytosine as indicated (\*) in the recognition sequence.

Storage buffer

20 mM Tris-HCl, 50 mM KCl, 0.1 mM EDTA 10 mM dithioerythritol, 0.01% polydocanol (w/v), 50% glycerol (v/v), pH 7.5 (at 4°C).

**Suppl. Incubation** buffer (10×)

330 mM Tris-acetate, 660 mM K-acetate, 100 mM Mg-acetate, 5 mM dithiothreitol, pH 7.9 (at 37°C), (△ SuRE/Cut Buffer A).

**Activity** in SuRE/Cut Buffer System

Bold face printed buffer indicates the recommended buffer for optimal activity:

Ī	Α	В	L	М	Н
ſ	100%	50-75%	25-50%	25-50%	0-10%

#### Incubation temperature

## 37°C

## **Unit definition**

One unit is the enzyme activity that completely cleaves 1 μg λDNA in 1 h at 37°C in a total volume of 25 μl SuRE/Cut Buffer A.

#### **Typical** experiment

Component	Final concentration
DNA	1 μg
10× SuRE/Cut Buffer A	2.5 μl
Sterile redist. water	Up to a total volume of 25 μl
Restriction enzyme	1 unit

Incubate at 37°C for 1 h.

## **Heat Inactivation**

The enzyme can be heat-inactivated by heating to 65°C for 15 min.

## Number of cleavage sites on different DNAs (2):

λ	Ad2	SV40	Φ X174	M13mp7	pBR322	pBR328	pUC18
143	158	34	24	24	16	14	1

#### **Activity** in PCR buffer

Relative activity in PCR mix (Tag DNA Polymerase buffer) is 100%. The PCR mix contained lambda target DNA, primers,10 mM Tris-HCl (pH 8.3, 20°C), 50 mM KCl, 1.5 mM MgCl<sub>2</sub>, 200 µM dNTPs, 2.5 U Taq DNA Polymerase. The mix was subjected to 25 amplification

#### Ligation and recutting assay

Alu I fragments obtained by complete digestion of 1 µg pBR322 DNA are ligated with 0.1 units T4-DNA ligase in a volume of 10  $\mu$ l by incubation for 16 h at 25°C in 66 mM Tris-HCl, 5 mM MgCl<sub>2</sub>, 1 mM dithioerythritol, 1 mM ATP, pH 7.5 (at 20°C) resulting in >80 % recovery of 1 µg pBR322 DNA fragments. Subsequent re-cutting with Alu I yields >95% of the typical pattern of pBR322 DNA × Alu I fragments.

#### **Troubleshooting**

A critical component is the DNA substrate Many compounds used in the isolation of DNA e.g., phenol, chloroform, EtOH, SDS, high levels of NaCl, metals (e.g., Hg<sup>2+</sup>, Mn<sup>2+</sup>) inhibit or alter recognition specificity of many restriction enzymes. Such compounds should be removed by ethanol precipitation followed by drying, before the DNA is added to the restriction digest reaction. Appropriate mixing of the enzyme is recommended.

#### **Quality control**

Lot-specific certificates of analysis are available at www.lifescience.roche.com/certificates.

#### Absence of unspecific endonuclease activities

1 μg λDNA is incubated for 16 h in 50 μl SuRE/Cut buffer A with excess of Alu I. The number of enzyme units which do not change the enzyme-specific pattern is stated in the certificate of analysis.

#### Absence of exonuclease activities

Approx. 5 g [3H] labeled calf thymus DNA are incubated with 3  $\mu$ l A/ $\mu$ l for 4 h at 37 °C in a total volume of 100  $\mu$ l, 50 mM Tris-HCl, 10 mM MgCl<sub>2</sub>, 1 mM dithioerythritol, pH approx. 7.5. Under these conditions, no release of radioactivity is detectable, as stated in the certificate of analysis...

#### References

- Roberts, R. J. et al. (1976) J. Mol. Biol. 102, 157 Kessler, C. & Manta, V. (1990) Gene 92, 1–248.
- Rebase The Restriction Enzyme Database: http://rebase.neb.com.
- Benchmate: http://www.roche-applied-science.com/benchmate.

## **Ordering Information**

Product	Application	Packsize	Cat. No.
Rapid DNA Liga- tion Kit	Ligation of sticky- or blunt-ended DNA fragments in just 5 min at +15 to +25 °C.	Kit (40 DNA ligations)	11 635 379 001
T4 DNA Ligase	Ligation of sticky- and blunt- ended DNA fragments.	100 U 500 units (1 U/μl)	10 481 220 001 10 716 359 001
rAPid Phosphatase	Dephosphorylation of 5'-phosphate residues from nucleic acids	1000 U 5000 U	04 898 133 001 04 898 141 001
rAPid Dephos and Ligation Kit	Dephosphorylation of nucleic acids.	40 reactions 160 reactions	04 898 117 001 04 898 125 001
Alkaline Phospha- tase (AP), special quality for molecu- lar biology	Dephosphorylation of 5'-phosphate residues from nucleic acids.	1000 U (20 U/µl)	11 097 075 001
Agarose MP	Multipurpose agarose for analytical and prepara- tive electrophoresis of nucleic acids	100 g 500 g	11 388 983 001 11 388 991 001
Agarose LE	Separation of nucleic acids in the range 0.2 - 1.5 kbp	100 g 500 g	11 685 660 001 11 685 678 001
Agarose Gel DNA Extraction Kit	For the elution of DNA fragments from agarose gels.	1 Kit (max. 100 reactions)	11 696 505 001
High Pure PCR Product Purifica- tion Kit	Purification of PCR or enzymatic modification reaction (e.g. restriction digest)	50 purifications 250 purifications	11 732 668 001 11 732 676 001
SuRE/Cut Buffer Set for Restriction Enzymes	Incubation buffers A, B, L, M and H for restriction enzymes	1 ml each (10× conc. solutions)	11 082 035 001
SuRE/Cut Buffer A	Restriction enzyme incubation	$5 \times 1$ ml ( $10 \times$ conc. solution)	11 417 959 001
SuRE/Cut Buffer B	Restriction enzyme incubation	$5 \times 1$ ml ( $10 \times$ conc. solution)	11 417 967 001
SuRE/Cut Buffer H	Restriction enzyme incubation	$5 \times 1$ ml ( $10 \times$ conc. solution)	11 417 991 001
SuRE/Cut Buffer L	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 975 001
SuRE/Cut Buffer M	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 983 001
Water, PCR Grade	Specially purified, double-distilled, deionized, and autoclaved	100 ml (4 vials of 25 ml) 25 ml (25 vials of 1 ml) 25 ml (1 vial of 25 ml)	03 315 843 001 03 315 932 001 03 315 959 001
BSA, special qual- ity for molecular biology	Maintaining enzyme stability	20 mg (1 ml)	10 711 454 001

## Changes to previous version

editorial changes

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## **Commonly used bacterial strains**

Strain	Genotype
BL21	E. coli B F $^-$ dcm ompT hsdS( $r_B$ - $m_B$ -) gal (Studier, F.W. et al (1986) J. Mol. Biol., <b>189</b> , 113.)
C600 <sup>e</sup>	supE44 hsdR2 thi-1 thr-1 leuB6 lacY1 tonA21; (Hanahan, D. (1983) <i>J. Mol. Biol.</i> <b>166</b> , 557.)
DH5α	supE44 Δ(JacU169 (φ80d/acZΔM15) hsdR17 recA1 endA1 gyrA96 thi-1 relA1; (Hanahan, D. (1983) J. Mol. Biol. <b>166</b> , 557.)
HB101	supE44 hsdS20 recA13 ara-14 proA2 lacY1 galK2 rpsL20 xyl-5 mtl-1; (Hanahan, D., (1983) J. Mol. Biol. 166, 557.)
JM108	recA1 supE44 endA1 hsdR17 gyrA96 relA1 thi $\Delta$ (lac-proAB); (Yanisch- Perron, C. et al., (1985) Gene <b>33</b> , 103.)
JM109	recA1 supE44 endA1 hsdR17 gyrA96 relA1 thi $\Delta$ (lac-proAB) F'[traD36proAB <sup>+</sup> , lacl <sup>q</sup> lacZ $\Delta$ M15]; (Yanisch- Perron, C. et al., (1985) Gene <b>33</b> , 103.)
JM110	rpsL (Str <sup>f</sup> ) thr leu thi-I lacY galK galT ara tonA tsx dam dcm supE44 Δ(lac-proAB) F[traD36proAB <sup>+</sup> , lacf <sup>f</sup> lacZΔM15]; (Yanisch- Perron, C. et al., (1985) Gene <b>33</b> , 103.)
K802	supE hsdR gal metB; (Raleigh, E. et al., (1986) Proc.Natl. Acad.Sci USA, 83, 9070.; Wood, W.B. (1966) J. Mol. Biol., <b>16</b> , 118.)
SURE <sup>r</sup>	recB recJ sbc C201 uvrC umuC::Tn5(kan <sup>r</sup> ) lac , Δ(hsdRMS) endA1 gyrA96 thi relA1 supE44 F[proAB <sup>+</sup> lacl <sup>q</sup> lacZΔM15 Tn10 (tet <sup>1</sup> ); (Greener, A. (1990) Stratagies, <b>3</b> , 5.)
TG1	supE hsd Δ5 thi Δ(lac-proAB) F <sup>*</sup> [traD36proAB <sup>+</sup> , lacl <sup>q</sup> lacZΔM15]; (Gibson, T.J. (1984) PhD Theses. Cambridge University, U.K.)
XL1-Blue <sup>r</sup>	supE44 hsdR17 recA1 endA1 gyrA46 thi relA1 lac F'[proAB $^+$ , lacl $^q$ lacZ $\Delta$ M15 Tn10 (tet $^D$ ]; (Bullock et al., (1987) BioTechniques, 5, 376.)

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