# ProductInformation

**Precast Mini-Polyacrylamide Gels**
Product Nos. P4966, P5091, P5216, P5341, P5466, P0473, P5591, P4841, P7341, P7466, P7591, P7716, P7841, P7966, P9347, P8091

**Caution: Packages contain 0.02% sodium azide in gel buffer**
Store flat at 2-8°C; DO NOT FREEZE

## Gel Descriptions

Cassette Sizes: 8 cm tall x 10 cm wide x 5 mm thick, 10 cm tall x 10 cm wide x 5 mm thick
Gel Sizes: 6.6 cm tall x 8.1 cm wide x 1 mm thick, 8.6 cm tall x 8.1 cm wide x 1 mm thick
Sample Format: All gels are cast with twelve wells that will each accommodate up to 20 µl sample volume.

## Compatibility

| Unit Manufacturer | 8 x 10 cm | 10 x 10 cm |
|---|---|---|
| Sigma-Aldrich Mini Techware | YES | YES |
| Bio-Rad Mini-Protean II[1] | YES | NO |
| Daiichi 2 & 6 Gel Systems | YES | YES |
| Hoefer Mighty Small (SE260) | NO | YES |
| Hoefer Mighty Small (SE250) | YES | NO |
| ISS/Enprotech | YES | YES |
| Life Technologies | YES | YES |
| Novex | YES | YES |
| Owl Scientific Penguin | NO | YES |
| Stratagene | YES | YES |

[1]Reverse device gasket matching flat side to gel

## Product Specifications

| Polyacrylamide Concentration | Prod. No. 8 x 10 cm | Prod. No. 10 x 10 cm |
|---|---|---|
| Tris-HCl buffer system (0.375M Tris-HCl, pH 8.9): Suitable for separating proteins and nucleic acids. Compatible with Tris-glycine-SDS running buffer for denatured proteins, Tris glycine for native proteins, and TAE or TBE for nucleic acids. Acrylamide/bis-acrylamide ratio 37.5:1. Recommended running voltage: 125V constant | | |
| 5% | ——— | P4841 |
| 7.5% | P4966 | P7341 |
| 10% | P5091 | P7466 |
| 12.5% | P5216 | P7591 |
| 15% | P5341 | P7716 |
| 4-20% | P5466 | P7841 |
| 10-18% | P0473 | P9347 |
| 10-27% | P5591 | P7966 |
| Tris-tricine buffer system: (1.0M Tris-HCl, pH 8.45): Suitable for separating low molecular weight proteins and peptides. Compatible with Tris-tricine-SDS cathode buffer and Tris-HCl anode buffer. Acrylamide/bis-acrylamide ratio 29:1. Recommended running voltage - 100V constant. | | |
| 10-20% | ——— | P8091 |

**Procedure**

**Caution: Gloves should always be worn when handling all polyacrylamide gels.**

1. Cut open the package and carefully remove the gel.
2. Rinse the gel with deionized water.
3. Remove the tape from the cassette bottom exposing the lower gel.
4. Grasp the ring on the comb, and gently pull the well-comb from the gel cassette.
5. Gently rinse the sample wells with deionized water and shake the water out.
6. Mount the gel onto the electrophoresis device with the front plate (well side) facing the inner cathode buffer chamber.
7. Fill the upper and lower chambers with the appropriate buffers and dislodge any air bubbles trapped under the gel bottom.
8. Load the samples. The wells will accommodate a maximum of 20 µl sample volume.
9. Attach the electrophoresis device to the power supply and run the gel using the suggested power settings until the dye front is near the gel bottom.
10. After the run is complete TURN OFF AND DISCONNECT THE POWER SUPPLY and remove the gel from the electrophoresis unit.
11. Separate the two cassette plates by inserting a spatula or any flat edged device into the recessed slot on the side, then twist to pry the cassette plates apart. Repeat the process along both sides of the cassette.
12. After separating the cassette plates, push on the bottom of the gel by running a spatula along the slot of the back plate.
13. Fix, stain and destain or transfer the gel as desired.

4/00